

NATURAL ENVIRONMENT RESEARCH COUNCIL









Initial results of long-term OA exposure on the biology of intertidal benthic foraminifera

Nikki Khanna (nk274@st-andrews.ac.uk)

Supervisors: Professor D. M. Paterson & Dr W. E. N. Austin University of St Andrews

In collaboration with Dr J. A. Godbold & Dr M. Solan University of Southampton

Funding for this project is provided by NERC and the University of St Andrews

Talk Outline:

- Introduction: Foraminifera and Ocean Acidification
- Details on Experimental System
- Sampling Regime and Foraminiferal Picking Methods
- Preliminary Results
- Future Work

1. Foraminifera & Ocean Acidification:

Changes in seawater chemistry due to ocean acidification will have severe implications on marine organisms that construct carbonate shells and structures, including foraminifera.

Foraminifera are unicellular organisms that construct a shell (test) which, when the animal dies can remain in the sediment as a fossil

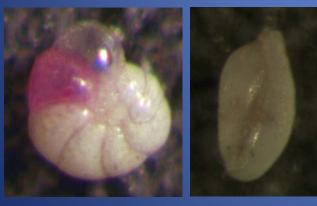
Shells (tests) hold the potential to record information on changing ocean chemistry, often leaving a microfossil record that permits reconstruction of environmental history



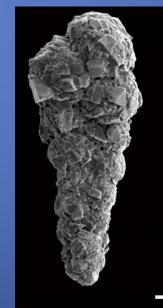
2. Foraminifera:

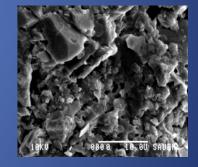
- Constitute one the most diverse groups of shelled microorganisms in modern oceans
- The type of shell material can determine where various species or their remains can survive
- Varying wall compositions = different sensitivities to dissolution

Calcareous forms secrete a CaCO₃ test



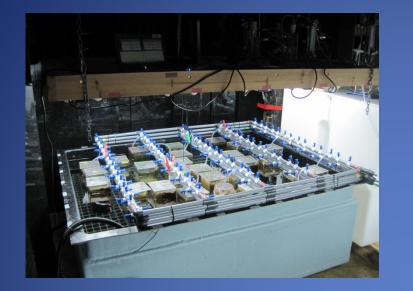
Hyaline Porcellaneous





Agglutinated forms build tests by cementing detrital material

3. Experimental System:





- Samples taken from long-term exposure experiments at Oceanlab, Aberdeen (now at University of Southampton)
- Mud sampled from Ythan Estuary, NE Scotland (Dec 2010)
- CO₂ treatments maintained by bubbling from a air-CO₂ mixing system
- Sub-samples from sediment cores to investigate the foraminiferal response over time

4. Sampling Regime:

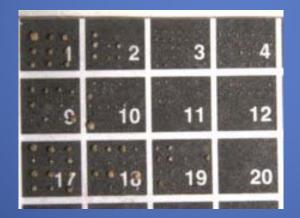
• Samples taken at 12 time points (January 2011 – March 2012)



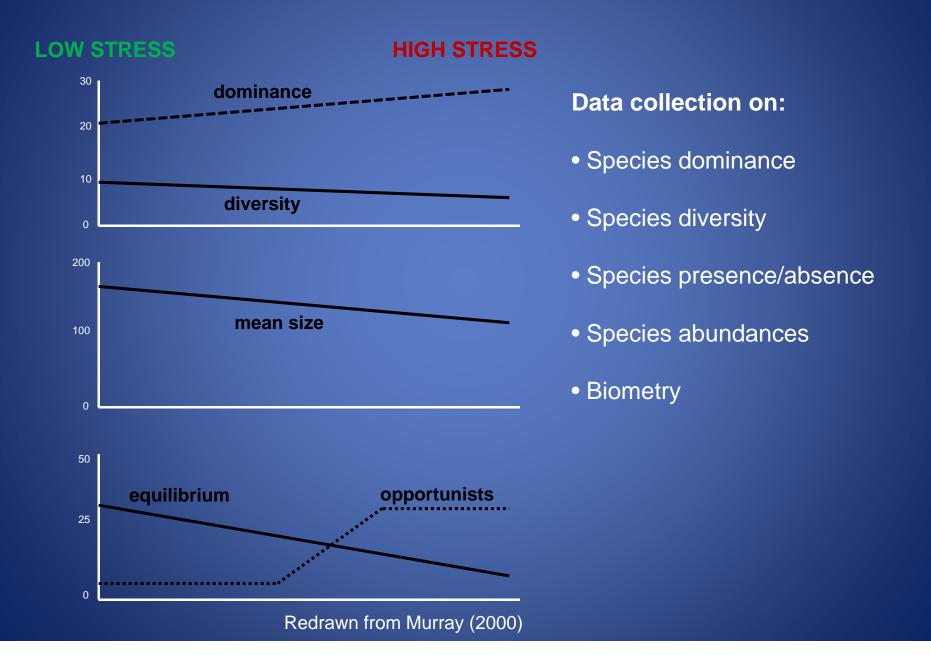
- CO₂ Treatments: 380, 750 and 1000 ppm
- Temperature: 10°C and Ambient
- Four replicates for each (combination of surface scrapes from three points within core)
- Same area never sampled twice

5. Sample Processing & Foraminiferal Picking Methods:

- Samples fixed and stained with Rose Bengal at time of sampling
- Sediment sieved (>63 μm), washed & dried (< 40 °C)
- Dry weights determined & samples stored
- Foraminifera extracted using standard micropalaeontological techniques



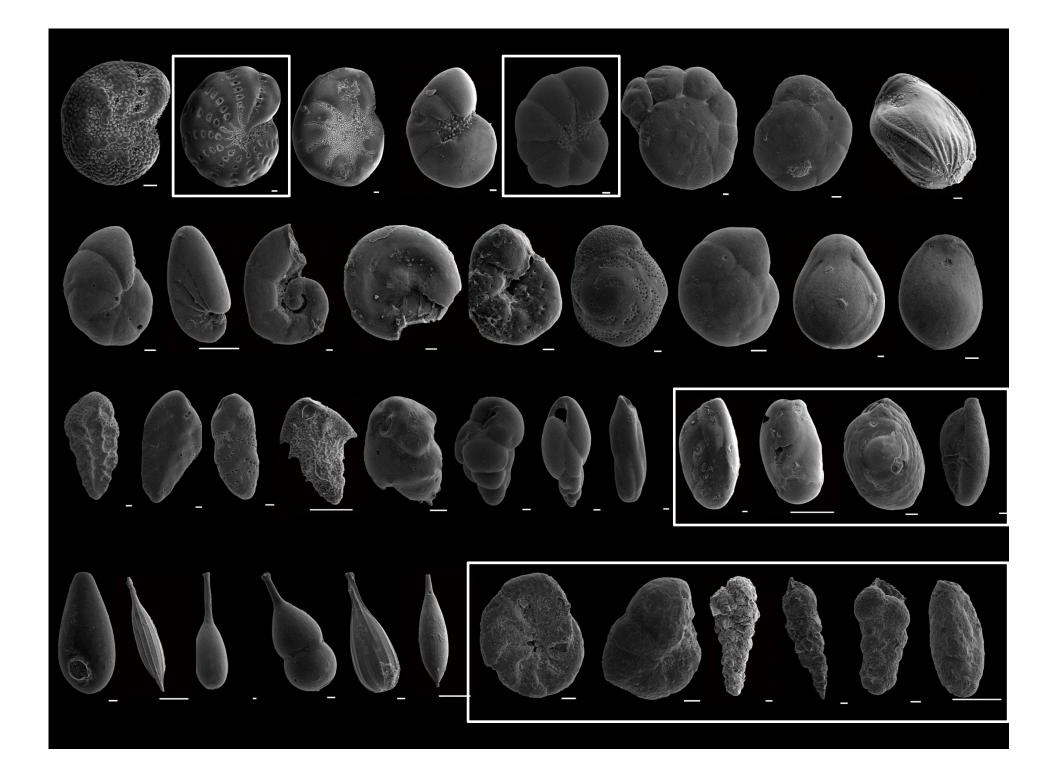
6. Predicted trends under environmental stress:



7. Preliminary Observations (1):

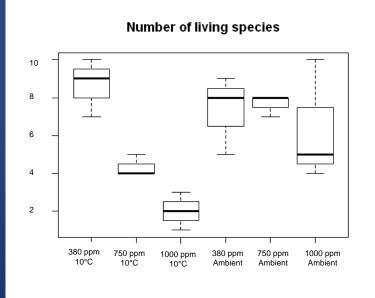
- 44 species identified from total (live + dead) assemblages
- Live foraminifera account for 1 22 % of total populations
- Two dominant species in live populations:
 - Haynesina germanica
 - Elphidium williamsoni *



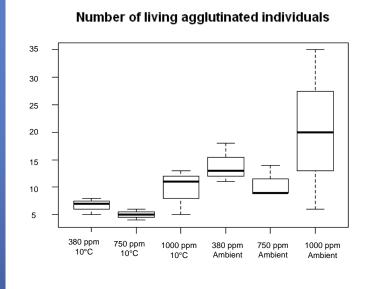


Preliminary Observations (2):

- Highest number of living species present at 380 ppm
- Fewer live individuals in highest CO₂ treatments
- More agglutinated individuals in highest CO₂ treatments





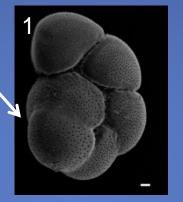


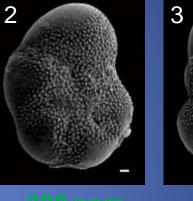
 $(df_1 = 2, df_{12} = 12, F = 4.411, p = 0.037)$

Preliminary Observations (3): Deformation and dissolution

Several different modes of abnormality in test morphology observed:

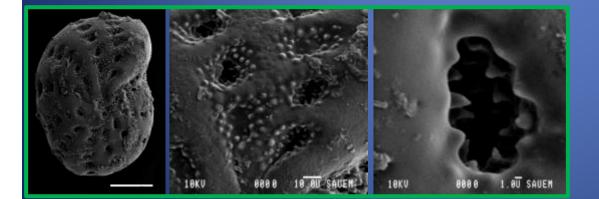
- Abnormal chamber additions
- Aberrant chamber shape
- Distorted chamber arrangement
- Reduced chamber size
- Reduction in ornamentation



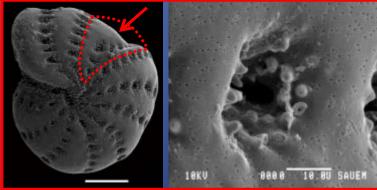


380 ppm -





380 ppm



1000 ppm

8. Future Work:

To provide valuable insight into foraminiferal response to ocean acidification:

- Keep picking forams! (Complete time-series)
- Establish a dissolution index
- Identify species specific sensitivities to changing pCO₂ & temperature
- Document changes in dominance and species diversity
- Quantify growth effects e.g. Maximum test diameter, test thickness

Thank you for your attention

