



UK Ocean Acidification
Research Programme
Benthic Acidification



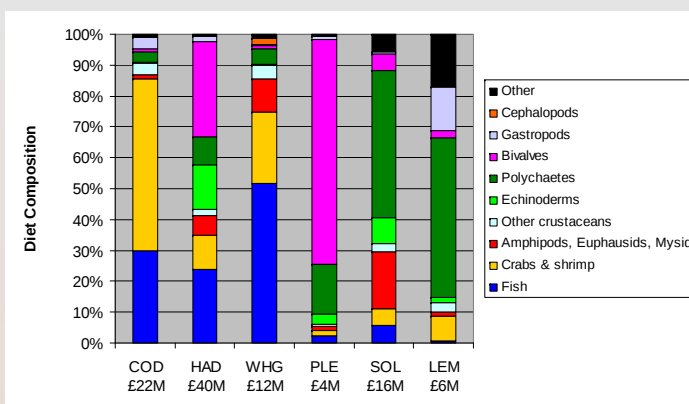
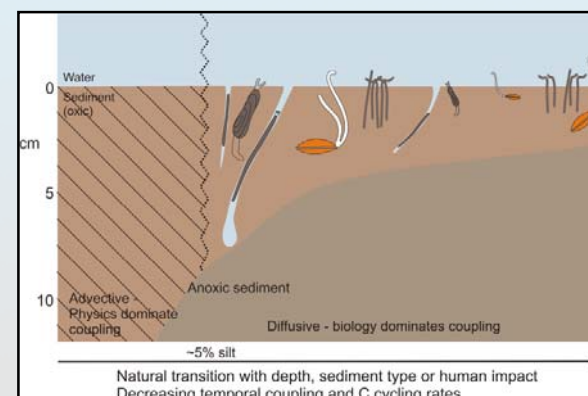
Scaling-up impacts to predict changes in ecosystem function

Silvana Birchenough, Ruth Parker, Julie Bremner, John Pinnegar & Finlay Scott



Monitoring changes in benthic communities...

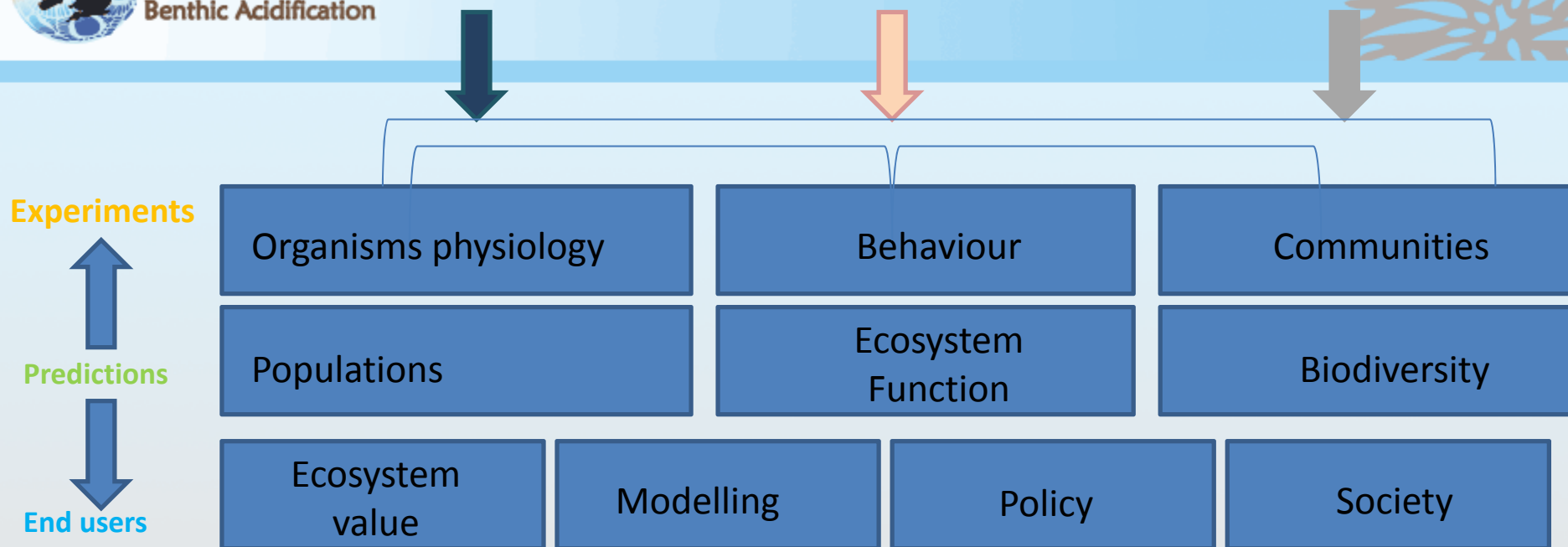
- Aim: to determine the effects of ocean acidification on the overall function of key benthic habitats
- To develop and test a conceptual framework to assess the broad-scale effects of OA in benthic communities (to 'scale-up' from experiments → assemblage → ecosystems)
- Results from high CO₂ environments, field and lab experiments will be used to inform the effect of OA on sediment functions
- Baseline assessment of species distribution and abundance (functional trait information)
- Functions: bioturbation, bioirrigation, production, provisioning for higher trophic levels



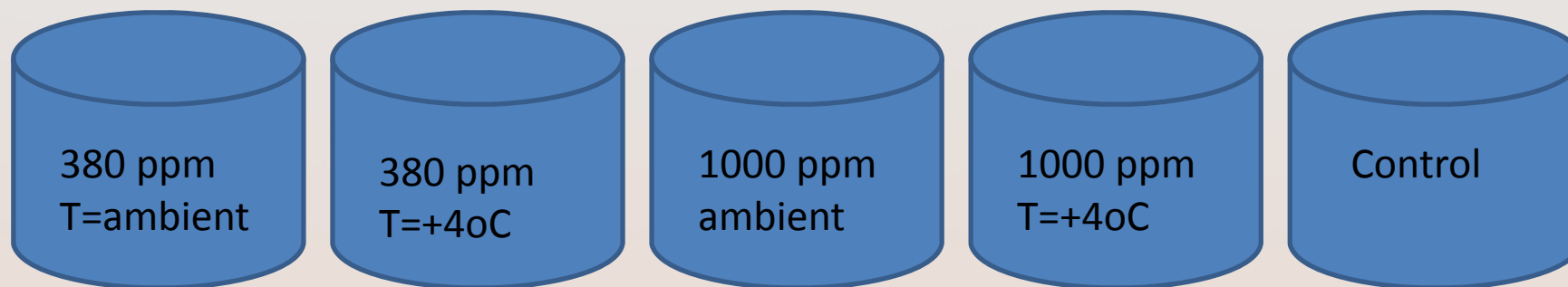


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Ocean Acidification



Amphiura filiformis , *Cerastodesma edule* and *Nereis virens*



Widdecombe, S., Dupont, S., Thorndyke. Laboratory experiments and benthic mesocosms studies. 2011. Guide to best practice for ocean acidification research and data reporting. Edited by U. Riebesell, Fabry, V.J., Hansson, L. Gattuso, J-P. Luxemburg, EU.

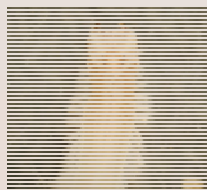
HYPOTHESIS



H₀: future high CO₂ scenarios will have no significant impact on the functioning of sediment habitats

Task: Map baseline species/assemblages=> functions and sediment biogeochemistry within the North Sea

- Benthic faunal work (abundance and BTA)
- Biogeochemical work (direct and indirect methods)
- Provisioning services – ‘bottom up’ effects on higher trophic levels (Fish)



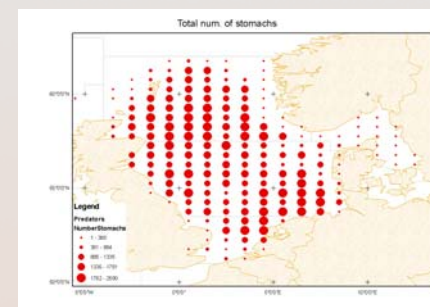
Data sources



- Identification of existing data sets (1950-1955, Kirkegaard, 1969), ICES North Sea Benthos (1986 and 2000), project surveys with information on species and their physical variables.
- Selection of traits to classify species /biological traits Analysis (BTA)
- Biogeochemical work (Defra funded projects E3205, E5301)
- Key functions based on key mediated biologically mediated ecosystem processes (bioturbation, bioirrigation, production)
- ICES and Cefas' DAPSTOM database for fish consumption=> in relation to the 3 benthic species (OA benthic consortium)

*The ICES dataset of fish stomach contents for 10 commercial species (good spatial coverage but limited number of predators)

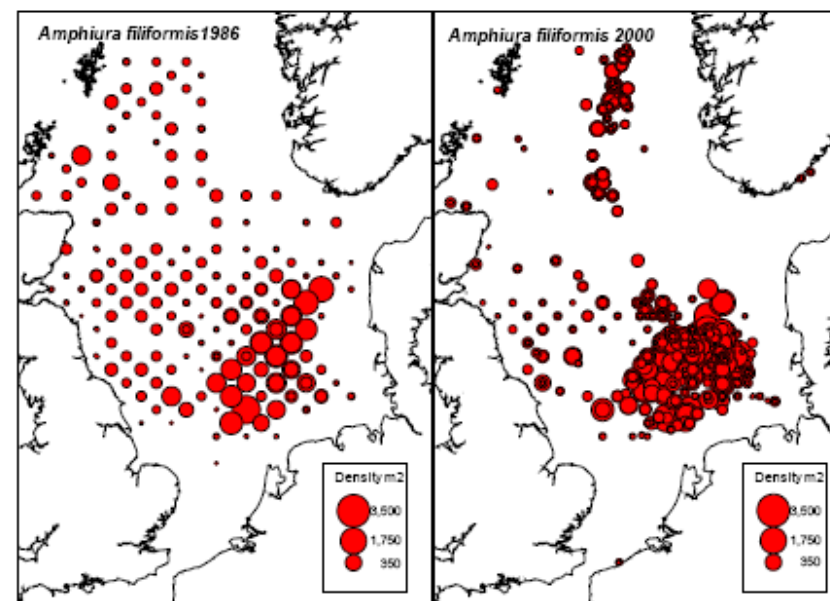
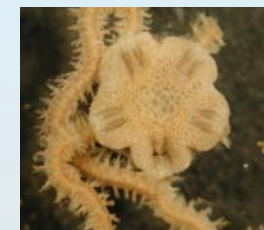
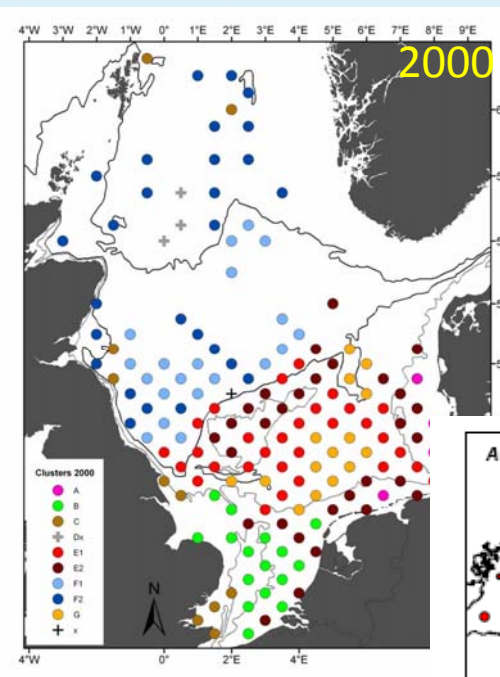
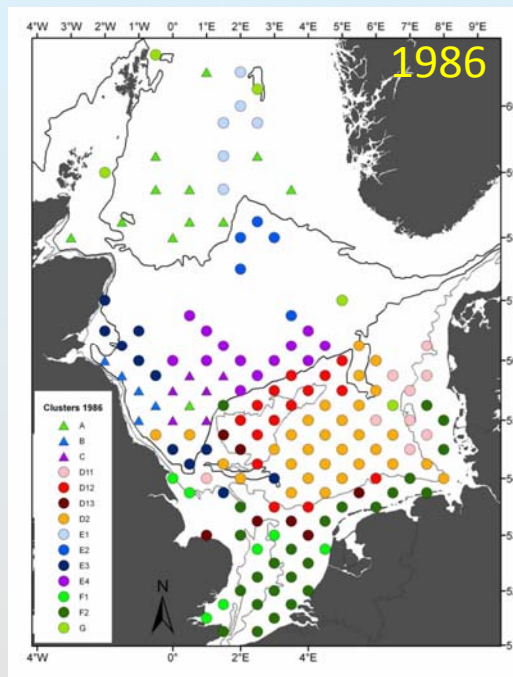
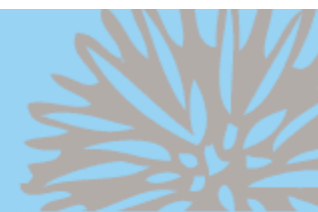
*The Cefas 'DAPSTOM' database that covers 149 different fish species, spanning the past 100 years (1893-2010) but limited spatial coverage





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Benthic assemblages



*Rees, H. L., et al , 2007, Kunitzer et al., 1992





Fauna



Species

Species traits

Assemblage traits

Species A



Deposit feeder
No migration
Burrower

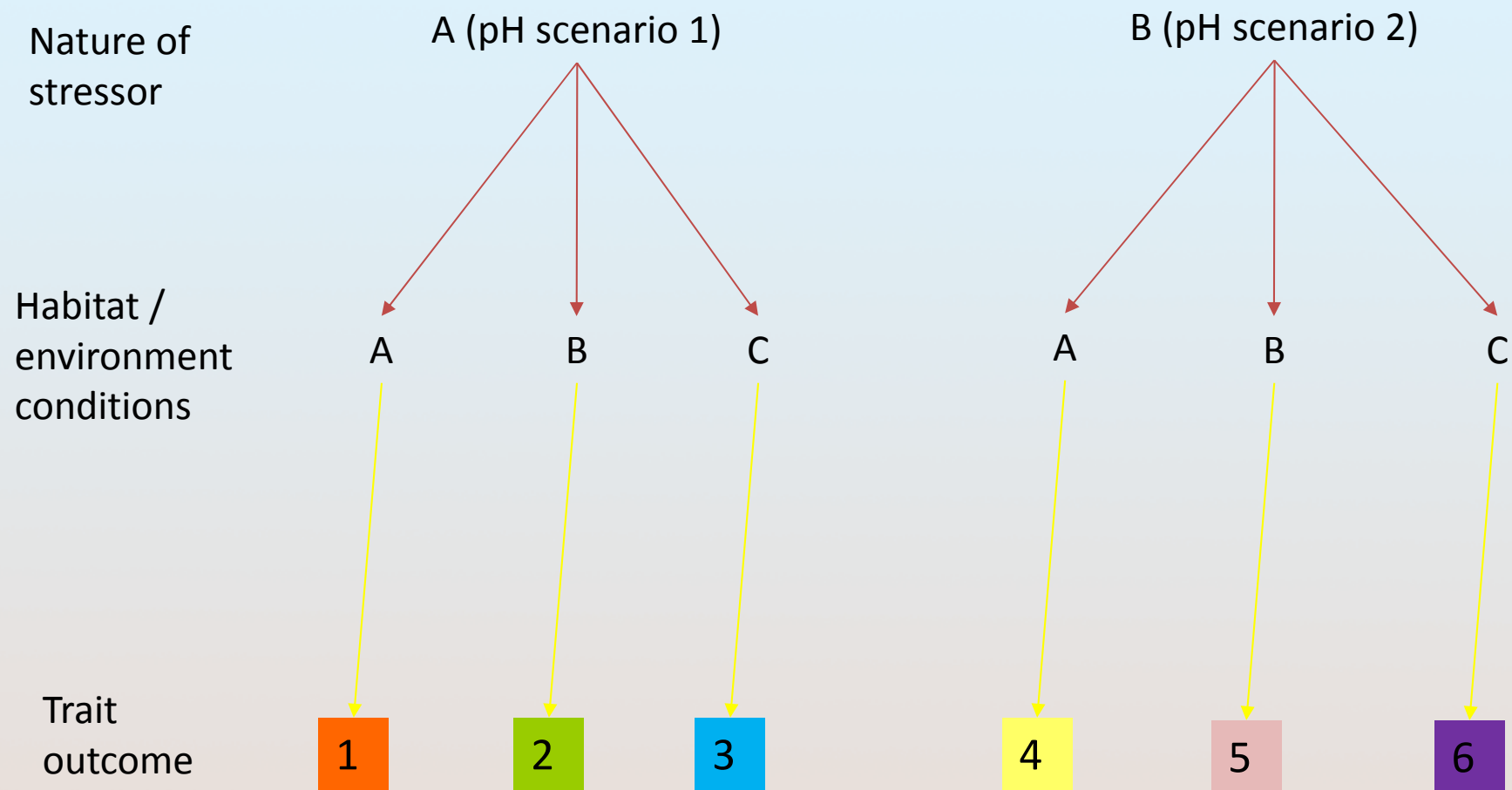
Species B



Scavenger
Horizontal migration
Vertical migration
Burrower
Crawler



Scavenger
Deposit feeder
No migration
Horizontal migration
Vertical migration
Burrower
Crawler





Biological traits



Trait	Function
Size	Carbon storage
Feeding	Energy transfer
Bioturbation	Nutrient cycling
Habit	Energy transfer
Reproduction	Connectivity export
Larval life	connectivity

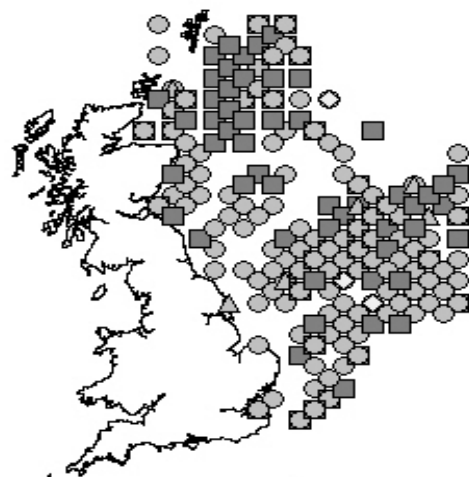
Energy

- Example of traits/attributes for benthic fauna
- Add traits to inform OA effects (e.g. shell, other responses)
- Species' sensitivities to OA
- A total of 935 taxa coded (fuzzy coding)
- Information available for most species / 'expert judgement'
- BTA analysis (Birchenough et al., 2011; Bremner et al., 2003a, Bremner et al., 2003b, 2006)

Traits	Attributes
Size (mm)	<20
	20-50
	50-100
	>100
Feeding	Carnivore (C)
	Scavenger (S)
	Selective deposit-feeder (sDF)
	Non-selective deposit-feeder (nsDF)
	Suspension-feeder (S)
Living habit	Tube
	Permanent burrow
	Free living
Bioturbation	Surface deposition (surf)
	Diffusive mixing (diff)
	Conveyor belt transport (convey)
	Reverse conveyorbelt (reverse)
	No bioturbation (No)
Reproduction type	Asexual (asex)
	Sexual-A
	Sexual-B



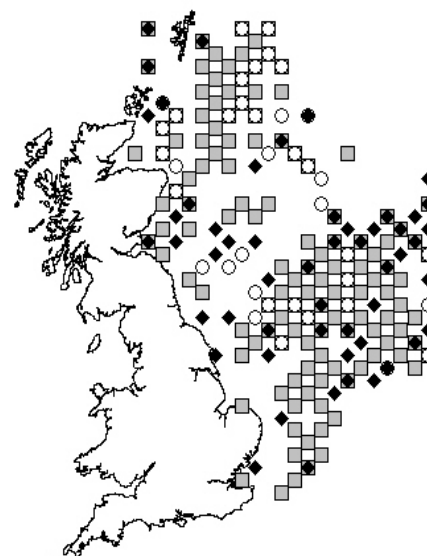
Patterns



Trophic groups

- 0.5=Surface feeder (144)
- 1=(ns) Deposit feeder (400)
- ◆ 2=Scavenger (26)
- ▲ 3=Carnivore (15)

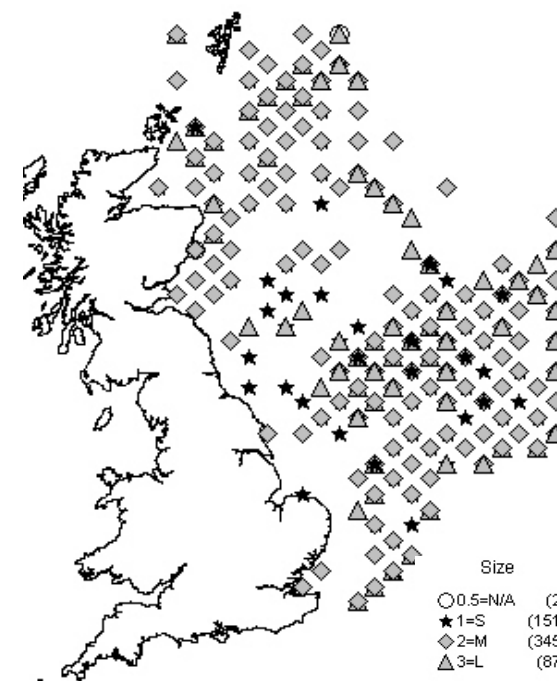
Feeding
Carbon Storage



Bioturbation

- 0.5=surt (298)
- 2=Conv (101)
- ◆ 3=biodiff (186)

Bioturbation
Nutrient cycling



Size

- 0.5=N/A (2)
- ★ 1=S (151)
- ◆ 2=M (345)
- ▲ 3=L (87)

Size
Energy transfer



Sensitivity review

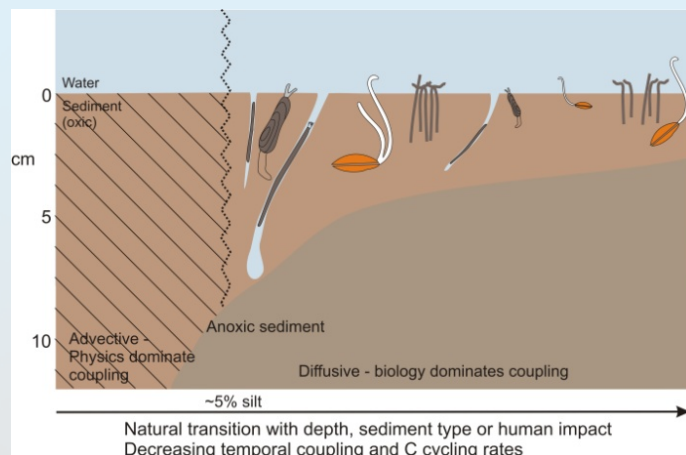


Taxa	Species	Description	CO ₂ system parameters	Sensitivity	Reference
Mollusca	<i>Haliotis laevis</i>	Greenlip abalone	pH 7.78; pH 7.39	5% and 50% growth reductions	Harris <i>et al.</i> (1999)
	<i>Haliotis rubra</i>	Blacklip abalone	pH 7.93; pH 7.37	5% and 50% growth reductions	Harris <i>et al.</i> (1999)
	<i>Mytilus edulis</i>	Blue Mussel	pH 7.1 / 10 000 ppmv	Shell dissolution	Lindinger <i>et al.</i> (1984)
			pCO ₂ 740 ppmv	25% decrease in calcification rate	Gazeau <i>et al.</i> (2007)
			pH 6.6	100% mortality within 30 days	Bamber (1990)
	<i>Mytilus galloprovincialis</i>	Mediterranean	pH 7.3, ~ 5000 ppmv	Reduced metabolism, growth rate	Michaelidis <i>et al.</i> (2005)
	<i>Crassostrea gigas</i>	Pacific Oyster	pCO ₂ 740 ppmv	10% decrease in calcification rate	Gazeau <i>et al.</i> (2007)
			pH 6.0	100% mortality within 30 days	Bamber (1990)
	<i>Placopecten magellanicus</i>	Giant scallop	pH < 8.0	Decrease in fertilization and embryo development	Desrosiers <i>et al.</i> (1996)
	<i>Tivela stultorum</i>	Pismo clam	pH < 8.5	Decrease in fertilization rates	Alvarado-Alvarez <i>et al.</i> (1996)
Arthropoda	<i>Pinctada fucata martensii</i>	Japanese pearl oyster	pH 7.7	Shell dissolution, reduced growth	Reviewed in Knutzen (1981)
			pH .7.4	Increasing mortality	
	<i>Mercenaria mercenaria</i>	Clam	Ω _{arag} =0.3	juv. shell dissolution, leading to increased mortality	Green <i>et al.</i> (2004)
	<i>Strombus luhuanus</i>	Gastropod	pH 7.9	Survival rate significantly lower	Shirayama and Thornton (2005)
Arthropoda	<i>Cancer pagurus</i>	Edible crab	1% CO ₂ ~10 000 ppmv	Reduced thermal tolerance, aerobic scope	Metzger <i>et al.</i> (2007)
	<i>Porcelana platycheles</i>	Porcelain crab	pH 7.4	after 40 days no effect detected	Calosi <i>et al.</i> (2009)
	<i>Callinassa</i> sp.	Mud shrimp	pH 6.3	tolerant	Torres <i>et al.</i> (1977)
	<i>Necora puber</i>	Swimming crab	pH 6.16	100% mortality after 5 days	
	<i>Amphibalanus amphitrite</i>	barnacle	pH 7.4 – pH 8.2	Weakening of shell	McDonald <i>et al.</i> (2009)
Echinodermata	<i>Strongylocentrotus pupuratus</i>	Sea urchin	pH ~6.2–7.3	High sensitivity inferred from lacking of pH regulation cf. Burnett <i>et al.</i> (2002) and	
	<i>Psammechinus miliaris</i>	Sea urchin		passive buffering via test dissolution during emersion	Spicer (1995); Miles <i>et al.</i> (2007)
	<i>Hemicentrotus pulcherrimus</i>	Sea urchin	~500–10 000 ppmv	Decreased fertilization rates, impacts larval developments	Kurihara and Shirayama (2004)
	<i>Echinometra mathaei</i>	Sea urchin			
	<i>Cystechinus</i> sp.	Deep-sea urchin	pH 7.8	80% mortality under simulated CO ₂ sequestration	Barry <i>et al.</i> (2002)
	<i>Amphiura filiformis</i>	brittlestar			Wood <i>et al.</i> (2008)
Sipuncula	<i>Sipunculus nudus</i>	Peanut worm	1% CO ₂ , 10 000 ppmv	Metabolic suppression	Pörtner and Reipschläger (1996)
				Pronounced mortality in 7 week exposure	Langenbuch and Pörtner (2004)
Polychaeta	<i>Nereis virens</i>		pH 6.5	tolerant	Batten and Bamber (1996)
	<i>Nereis virens</i>				Widdecombe <i>et al.</i> (2007)
Nematoda	<i>Procepalotrix similis</i>		pH < 5.0	tolerant	Yanfang and Shichum (2005)
Foraminifera	<i>Marginoptera kudakajimensis</i>		pH 7.7 – pH 8.3	Decline in calcification rate, possibly precluding survival	Kuroyanagi <i>et al.</i> (2009)



Modelling effects of OA on regional biogeochemical functioning: Methods

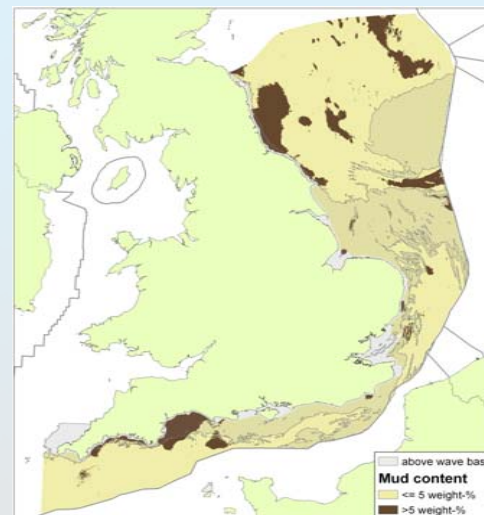
Regional distribution of biogeochemical status and controls



Direct: pH changes in water column propagation into the sediment – different sediment types (carbonate, quartz, shell ratios). Early stages – regional survey.

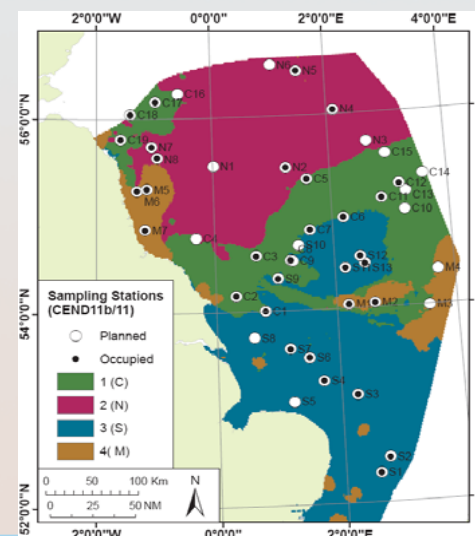
Indirect: Changes in macrofaunal assemblage (existing method)

Spatial and site specific modelling of existing datasets
Links statistical and ERSEM modelling



Areas where biogeochemical cycling processes are physically or biologically mediated. Permeable and not ~ wave base and % fines.

From Diesing and Parker, 2011 – Defra E5301



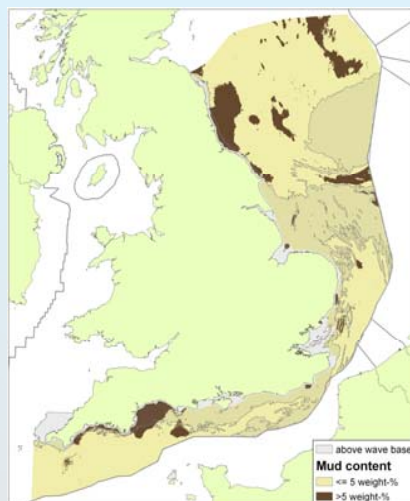
2011
Sampling of pH profiling and Sediments (seabed integrity)

*Greenwood et al. 2012

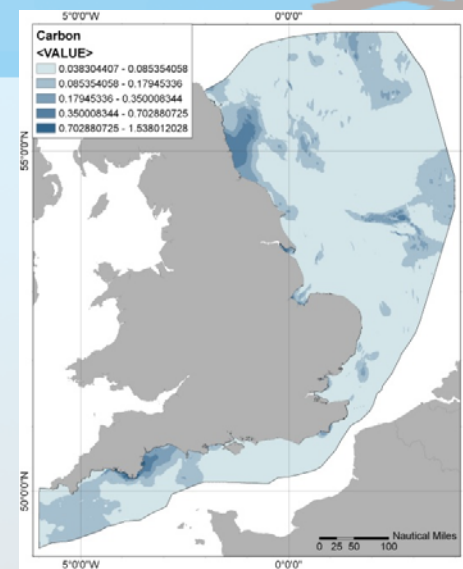
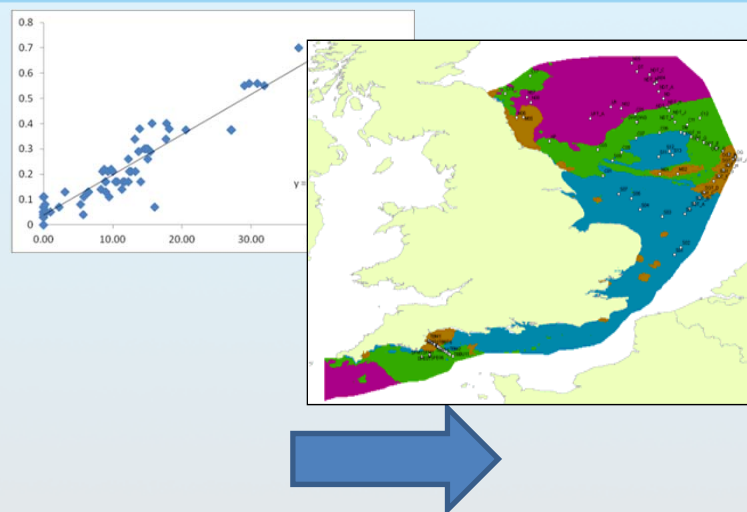


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Biogeochemical Baselines



% silt/clay



Carbon (%)

Biogeochemical status – North Sea

OPD

Carbon and Pigments

Nutrients

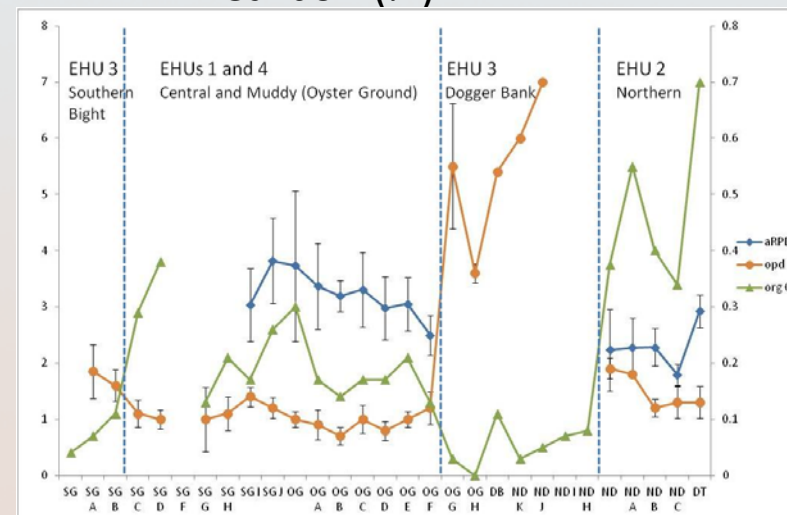
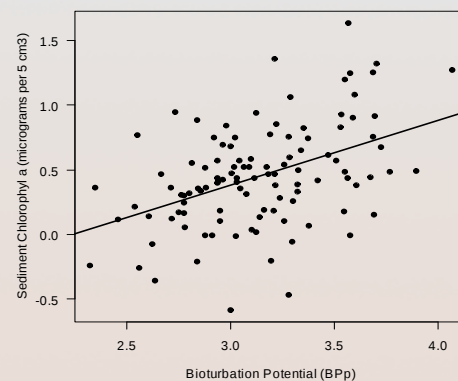
Redox (aRPD)

Linking to

Biological metrics

Production / P:B

Bioturbation (BPC)





Provisioning services – ‘bottom up’ effects on higher trophic levels

- Assessing the consumption by fish on specific benthic invertebrate species that are thought to be vulnerable to ocean acidification

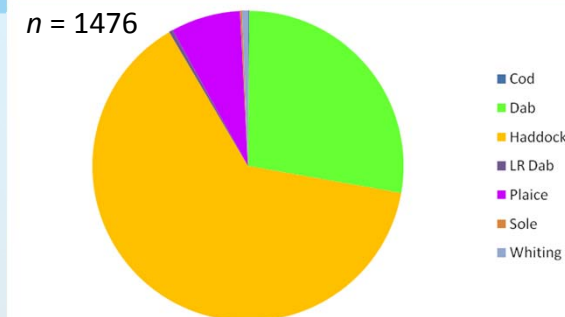
The ‘DAPSTOM’ dataset

The DAPSTOM database now contains 177,827 records from 157,159 individual predator stomachs and 149 fish species.

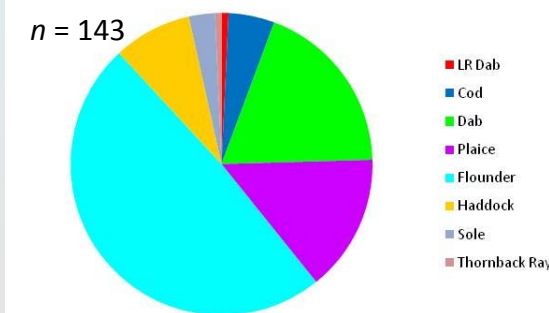
The full dataset is searchable via an online data portal (www.cefas.defra.gov.uk)

The dataset can be searched by “predator” species (e.g. haddock) or “prey” species (e.g. *Amphiura*).

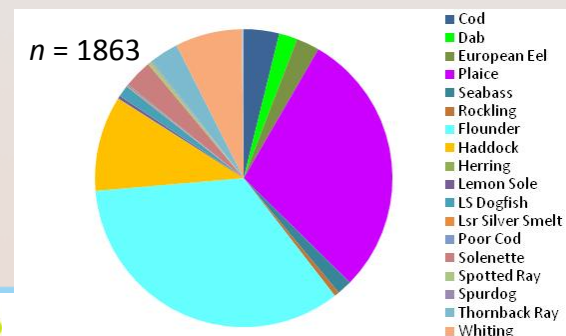
Predators feeding on *Amphiura filiformis* (% of records in database)

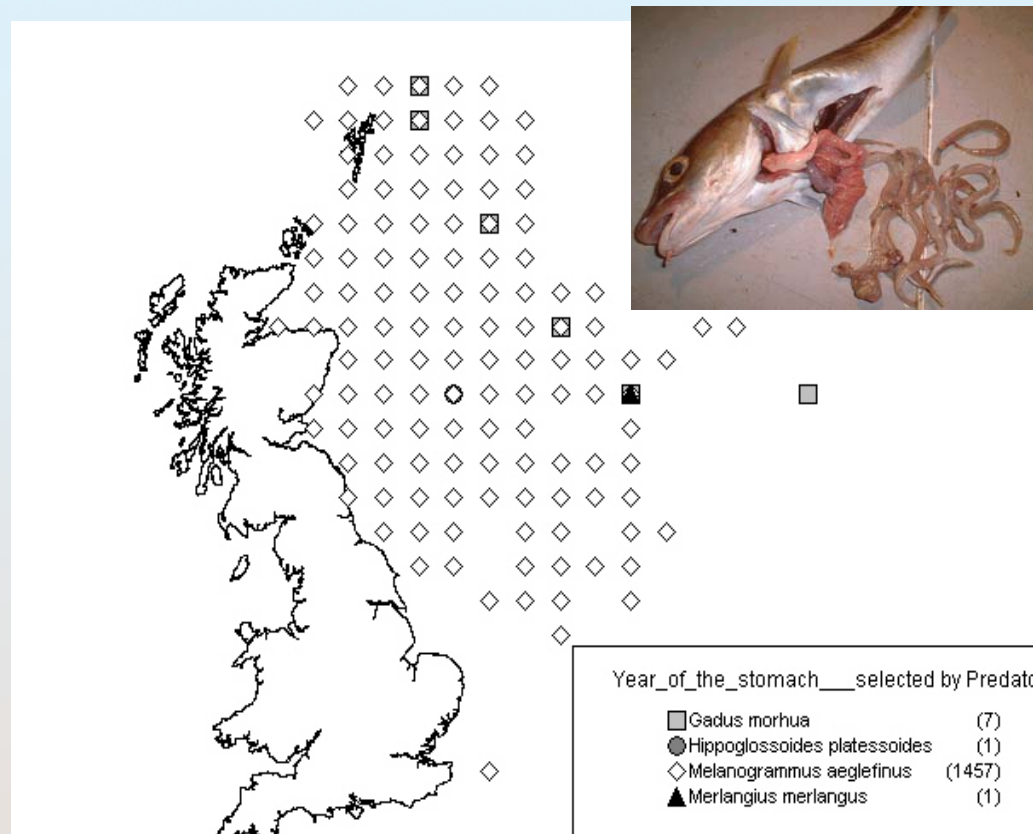
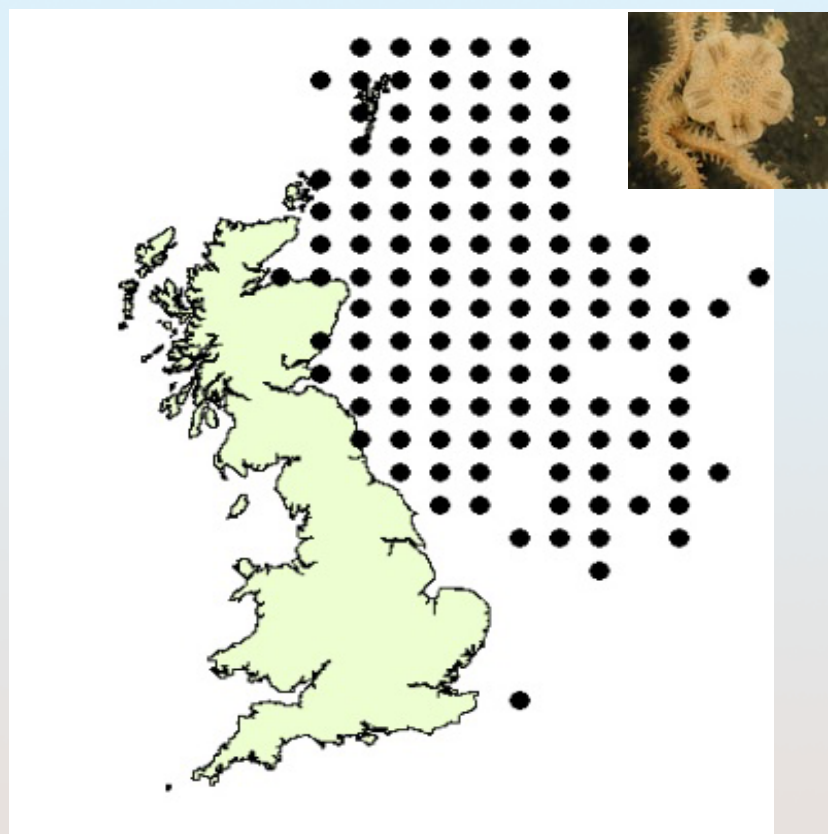


Predators feeding on *Cerastodesma edule*



Predators feeding on *Nereis/Nephtys* spp.

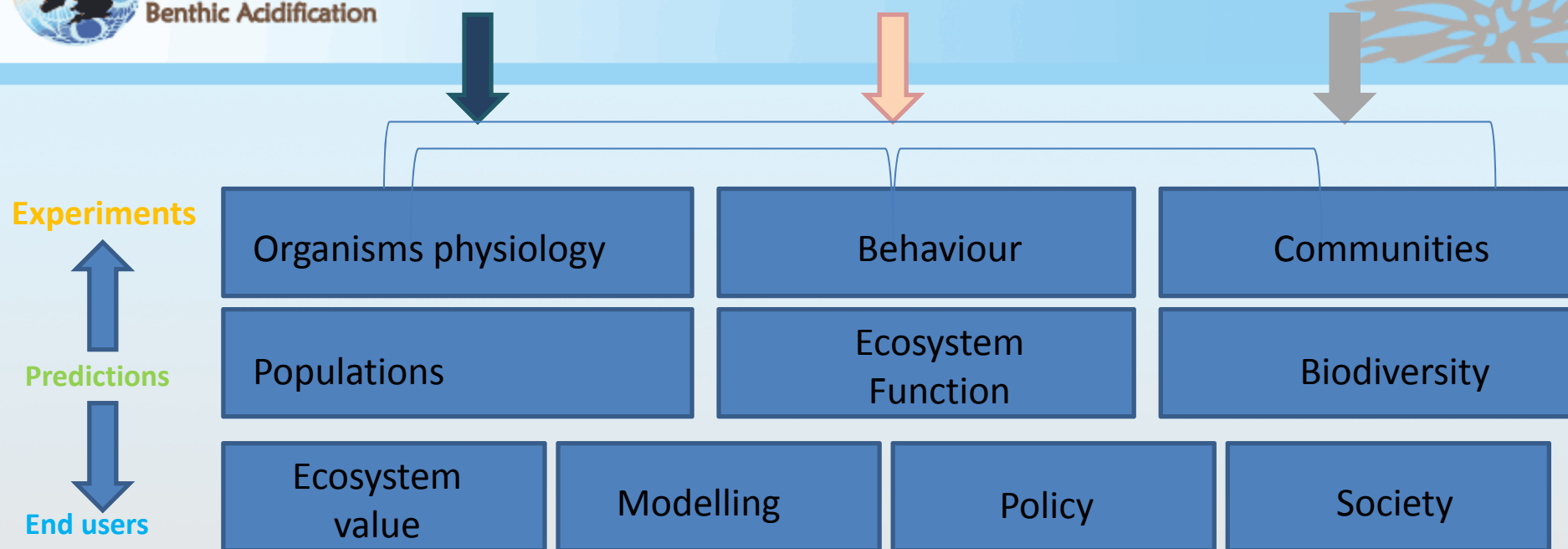




- Main predators (Haddock, Cod, Plaice, Whiting)
- Physiological effects on fish (need more information)
- How different fish predators might respond in the future to the changing patterns of prey availability (Kaplan et al., 2010)



Ocean Acidification



- This approach is a direct method to assess functional responses to 'OA' covering a number of benthic assemblages/sediment types in the NS.
- This information is complementary to experimental evidence and will help us to identify possible OA impact/areas
- This work will be used to 'scale up' from to possible impacts on ecosystem function (e.g. bioturbation, bioirrigation, production, nutrient cycling, provisioning for higher trophic levels)



Questions?



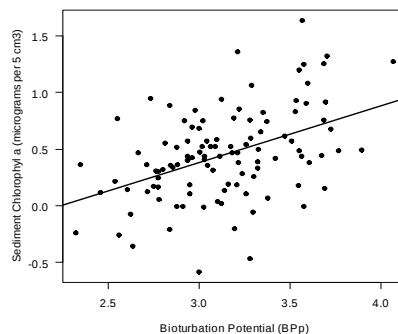


BPC vs sediment biogeochemistry or pH change ~
carbonate/shell etc

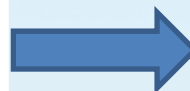


Biogeochemical modelling:

Coincident
Biogeochemical &
Faunal data
Site specific or
regional



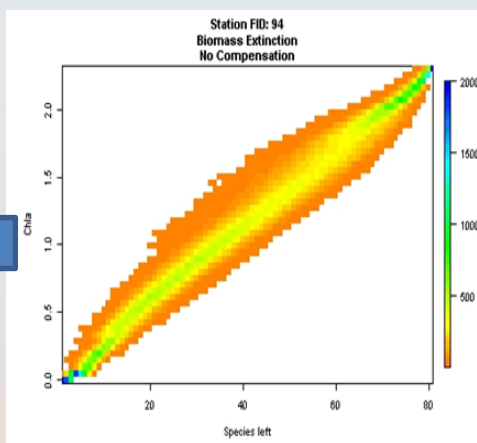
Sea region or
Sub-region



OA sensitivity
matrix of
species or
substrate type



Assemblage
species or
substrate altered
and BPC/function
recalculated



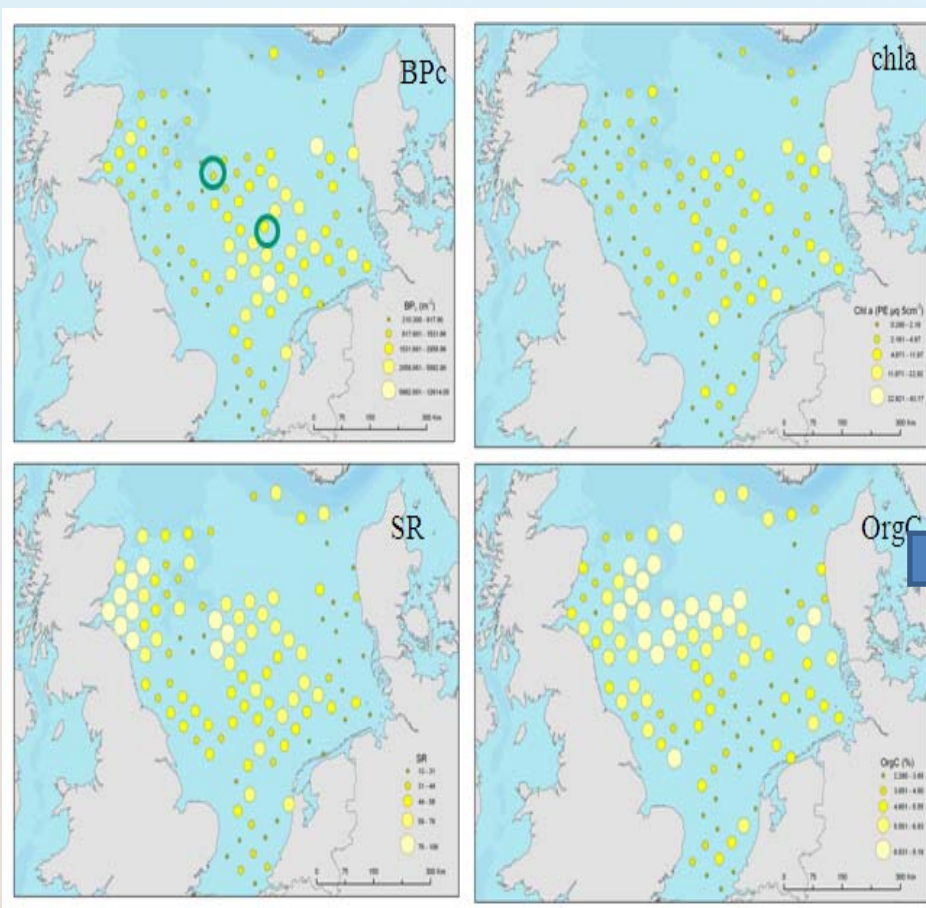
Biogeochemistry and BPC
tracked as species removed
or under set pH scenarios



Regional
futures or
rates of
change of
functions



Results: Baseline conditions- NSBS 1986

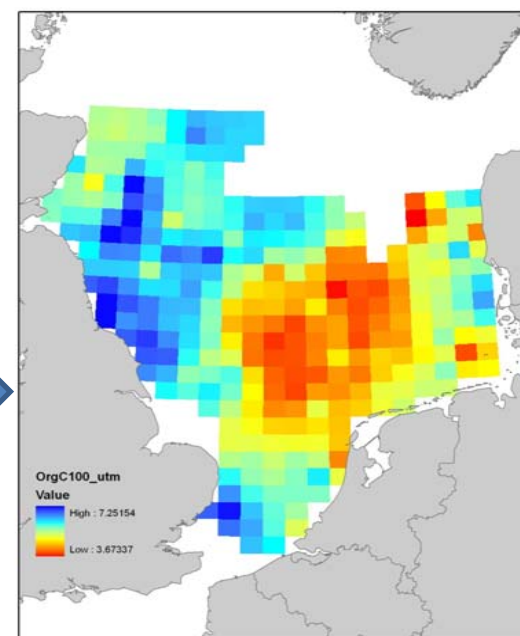


Observations:

Bioturbation (BPc, Solan et al., 2004)

Species richness

Carbon fractions



GIS mapping and volume calculations

Total regional budget as whole or part regions or changes given assemblage change.